

## Histologic Evaluation of an Nd:YAG Laser-Assisted New Attachment Procedure in Humans



Raymond A. Yukna, DMD, MS\*

Ronald L. Carr, DDS\*\*

Gerald H. Evans, DDS\*

*This report presents histologic results in humans following a laser-assisted new attachment procedure (LANAP) for the treatment of periodontal pockets. Six pairs of single-rooted teeth with moderate to advanced chronic periodontitis associated with subgingival calculus deposits were treated. A bur notch was placed within the pocket at the clinically and radiographically measured apical extent of calculus. All teeth were scaled and root planed with ultrasonic and hand scalers. One of each pair of teeth received treatment of the inner pocket wall with a free-running pulsed neodymium:yttrium-aluminum-garnet (Nd:YAG) laser to remove the pocket epithelium, and the test pockets were lased a second time to seal the pocket. After 3 months, all treated teeth were removed en bloc for histologic processing. LANAP-treated teeth exhibited greater probing depth reductions and clinical probing attachment level gains than the control teeth. All LANAP-treated specimens showed new cementum and new connective tissue attachment in and occasionally coronal to the notch, whereas five of the six control teeth had a long junctional epithelium with no evidence of new attachment or regeneration. There was no evidence of any adverse histologic changes around the LANAP specimens. These cases support the concept that LANAP can be associated with cementum-mediated new connective tissue attachment and apparent periodontal regeneration of diseased root surfaces in humans. (Int J Periodontics Restorative Dent 2007;27:577-587.)*

\*Professor, Department of Periodontics, Louisiana State University School of Dentistry, New Orleans, Louisiana.

\*\*Professor, Department of Oral Pathology, Louisiana State University School of Dentistry, New Orleans, Louisiana.

Correspondence to: Dr Raymond A. Yukna, Advanced Periodontal Therapies, University of Colorado Dental School, 13065 East 17th Place, Room 111, P.O. Box 6508, MS F847, Aurora, CO 80045; fax: 303-724-0162; e-mail: ray.yukna@uchsc.edu.

Regeneration of the supporting tissues of the teeth is a primary goal of periodontal therapy. Whereas clinical results and animal histology suggest that new connective tissue attachment (CTA) as well as regeneration of cementum (CEM), periodontal ligament (PDL), and alveolar bone (AB) can occur on human teeth affected by periodontitis as a result of several treatment approaches, histologic evidence in humans of successful cases and successful treatments is limited.<sup>1</sup>

The 1996 World Workshop in Periodontics established specific histologic criteria for proof of regeneration. Experimental teeth must have loss of CTA and AB associated with periodontitis. In addition, subgingival and/or subcrestal calculus must be present at the time of surgery so that a notch can be made into the root at the apical extent of calculus. Proof of new attachment is demonstrated by new CEM and CTA, and regeneration is evidenced by the presence of new CEM, PDL, and AB coronal to the apical extent of the notch. Most treatments that show proof of new attachment and regeneration are associated with surgically implanted devices or materials.<sup>1-21</sup>

Sulcular/pocket epithelium removal has been the basis or foundation of subgingival curettage (CUR), the excisional new attachment procedure (ENAP), and the replaced flap/modified Widman flap procedure to set up an environment for new CTA.<sup>22-25</sup> However, elimination of pocket epithelium by CUR, ENAP, or other internal-bevel incision designs appears nearly impossible.<sup>26</sup>

Procedures limited to treating the soft tissue wall of periodontal pockets such as CUR and ENAP would not be expected to influence new bone formation to any great degree but hopefully would lead to healing with a CTA rather than a long junctional epithelium (LJE). Almost all available human histologic evidence to date demonstrates healing by an LJE with no or minimal CTA.<sup>27</sup>

Interest in neodymium:yttrium-aluminum-garnet (Nd:YAG) laser use in periodontics is increasing. Several papers have suggested favorable results with its use in the treatment of periodontal pockets.<sup>28-30</sup> A procedure called laser ENAP has been promoted in trade journals with examples of radiographic bone regeneration.<sup>31,32</sup> Referred to as the laser-assisted new attachment procedure (LANAP) in this report, this technique of pocket therapy has recently been approved by the US Food and Drug Administration (FDA 510k clearance K030290).

In clinical case reports LANAP has demonstrated improved clinical measurements and some radiographic evidence of bone regeneration in the areas treated.<sup>33-37</sup> However, it is not known what tissues constitute the new healed interface between the soft tis-

ues and the tooth root. Also, there is some evidence that the use of lasers in periodontal pockets may damage root surfaces,<sup>38-49</sup> adversely affect the adjacent alveolar bone,<sup>50,51</sup> or cause undesirable pulpal changes.<sup>48,49</sup> Clinical case reports have reported favorable results, but there is no human histologic proof of the nature of the healing following LANAP. The purpose of this paper is to report histologic wound healing following use of LANAP surgery for periodontal pockets.

### Method and materials

Dental radiographs of patients assigned to the Postgraduate Periodontics Clinic, Louisiana State University (LSU), were screened for the presence of teeth that had isolated moderate to severe periodontal involvement (probing depths and clinical probing attachment loss of 5 to 9 mm with bleeding on probing and evident subgingival calculus). Teeth that had been treatment planned by clinicians in the Oral Diagnosis and/or Prosthodontics departments for extraction as part of the overall restorative treatment plan were included in the study. Subjects had to provide two single-rooted teeth with similar periodontal involvement for the study and signed an LSU-approved consent form prior to beginning the study.

Preoperatively, the subjects received occlusal adjustment/odontoplasty to reduce occlusal forces on the experimental teeth, and study teeth were splinted to neighboring teeth with an extracoronally bonded splint (Ribbond, Ribbond Inc). Scaling and

root planing were performed on other teeth in the same segment (not the treatment teeth), and general supragingival prophylaxis was provided for the rest of the mouth.

Documentation consisted of clinical photographs, radiographs with stent and grid (Fig 1), modified Gingival Index (mGI),<sup>52</sup> Quigley-Hein Plaque Index (PI),<sup>53</sup> and clinical mobility evaluation.<sup>54</sup> Clinical measurements were made from the cemento-enamel junction (CEJ) to the free gingival margin, from the CEJ to the base of the pocket, from the CEJ to the apical extent of clinically and radiographically evident calculus, and from the CEJ to the mucogingival junction. Bleeding on probing (BOP) was also assessed.

Appropriate laser safety precautions were used. Under regional local anesthesia, a quarter-round bur notch was placed at the clinically and radiographically measured apical extent of calculus as carefully as possible. One of each pair of teeth randomly received Nd:YAG laser treatment (Periolase, Millennium Dental Technologies) of the inner pocket wall to remove the crevicular epithelium around the necks of the study teeth, relax the gingival collar, and expose more of the contaminated root surface. The fiber tip of the laser was directed parallel to the root surface and was moved laterally and apically along the pocket wall, eventually reaching close to the base of the pocket. The laser settings for this first pass were 3 W, 150- $\mu$ s pulse duration, and 20 Hz. Once the epithelial lining was removed, root debridement was accomplished coronal to the area of the calculus reference notch with ultra-



**Fig 1** Radiographs of a 54-year-old man with deep infrabony defects on the mesial of both maxillary canines.

**Fig 1a** (left) Mesial defect on the right canine (left), treated with LANAP, demonstrates a radiographic increase in bone density and apparent fill of defect at 3 months after treatment (right).



**Fig 1b** (right) Mesial defect on the left canine (left), treated with scaling and root planing without laser, demonstrates little change in the bony defect contour or bone density after 3 months (right).

sonic (EMS Piezon 400, EMS) and hand instrumentation. No attempt was made to remove any soft/granulation tissue with the mechanical instrumentation. The pocket contents of the test teeth were lased again (4 W, 635- $\mu$ s pulse duration, and 20 Hz) to help achieve a solid fibrin clot and form a pocket seal. The control teeth received all of the aforementioned treatment except for the laser therapy. No sutures were used, and triple antibiotic ointment and a light-cured dressing (Baricaid, Dentsply/Caulk) were placed on all teeth. All patients were provided with nonsteroidal anti-inflammatory medications, doxycycline (100 mg daily for 10 days), and 0.12% chlorhexidine rinses (to be used twice daily).<sup>55</sup>

After 3 months, a second surgical procedure was performed to remove the experimental tooth roots en bloc according to methods described previously.<sup>4,9,56,57</sup> For all teeth this was a single proximal area. The body of each tooth root was bisected longitudinally in a faciolingual plane, with the clinician attempting to keep at least half of the

root diameter attached to the area of interest. A small interproximal wedge of tissue and a section of root approximately 5 mm wide, 7 mm long, and 5 mm thick was removed. Once the desired specimens were completely freed, they were gently and atraumatically removed, rinsed gently in sterile saline, and placed in 10% neutral buffered formalin. The residual defects were reconstructed, and after an appropriate healing period, the patients were referred for prosthetic replacements.

The biopsy specimens were processed by the LSU School of Dentistry Research Histology Laboratory, where they were decalcified, embedded in paraffin so as to obtain longitudinal mesiodistal serial step sections, serially sectioned at 7  $\mu$ m in the area of the notch, and stained with hematoxylin and eosin. The three most central 200- $\mu$ m serial step sections were blindly and randomly evaluated for the nature of the healed tissues—specifically the presence and length of new CEM, new

CTA, new AB, and healed junctional epithelium relative to the apical extent of the calculus notch. Histomorphometric measurements were made by an oral pathologist (RLC) using an eyepiece grid on the microscope. Root resorption, ankylosis, pulpal changes (where pulp tissue was visible), and the degree of inflammation were also evaluated. Mean values for the three sections of each tooth were used for linear measurements.

## Results

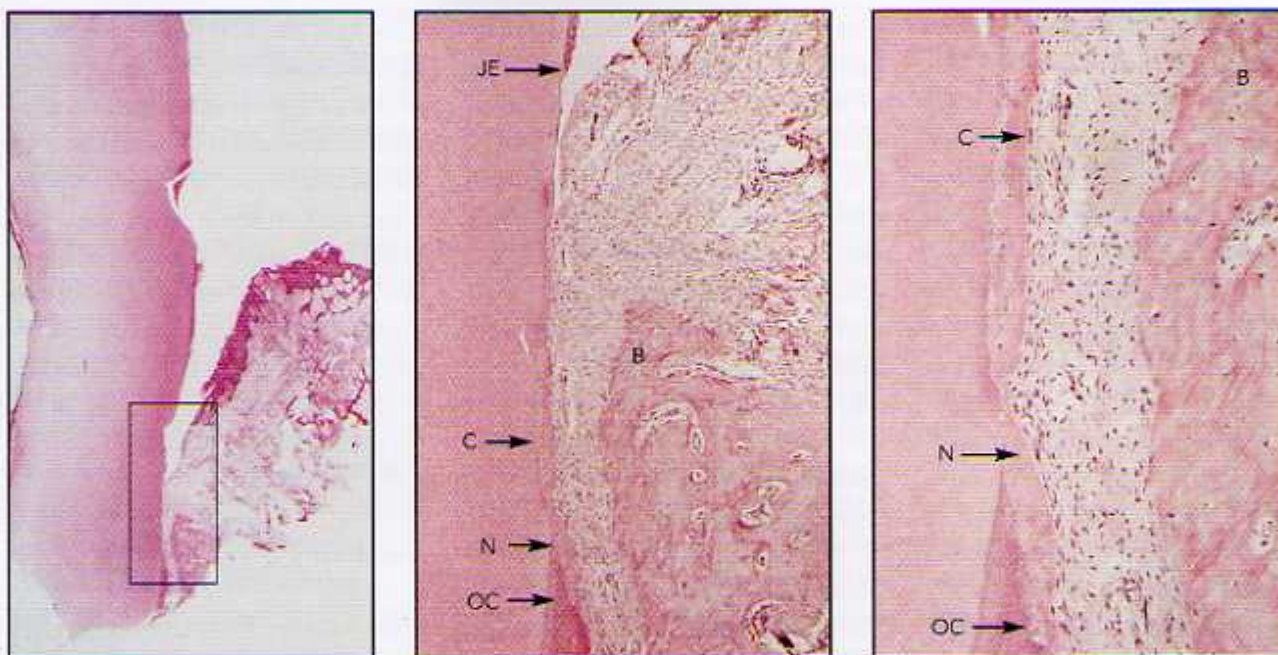
Three men and three women, 26 to 54 years old (mean 45.5 years), provided two teeth each. All subjects tolerated the treatment procedures well and reported that almost no pain-relieving medication was needed after the laser treatment. All teeth healed uneventfully. The LANAP-treated teeth exhibited greater mean probing depth reduction (4.7 mm vs 3.7 mm) and greater clinical probing attachment level gain (4.2 mm vs 2.4) than the con-

**Table 1** Clinical changes (3-month mean results, in millimeters) following use of LANAP or scaling and root planing alone (n = 6 teeth for each treatment)

Measurement/treatment	Pretreatment	3 mo	Change
Gingival recession			
LANAP	0.2*	0.1*	0.2
SCL/RP	0.3*	0.6	0.8
Probing depth			
LANAP	7.3	2.7	4.7
SCL/RP	8.0	4.3	3.7
Vertical CAL			
LANAP	7.2	3.0	4.2
SCL/RP	7.6	5.3	2.4

LANAP = laser-assisted new attachment procedure; SCL/RP = scaling and root planing; CAL = clinical attachment level.

\*Coronal to cemento-enamel junction.



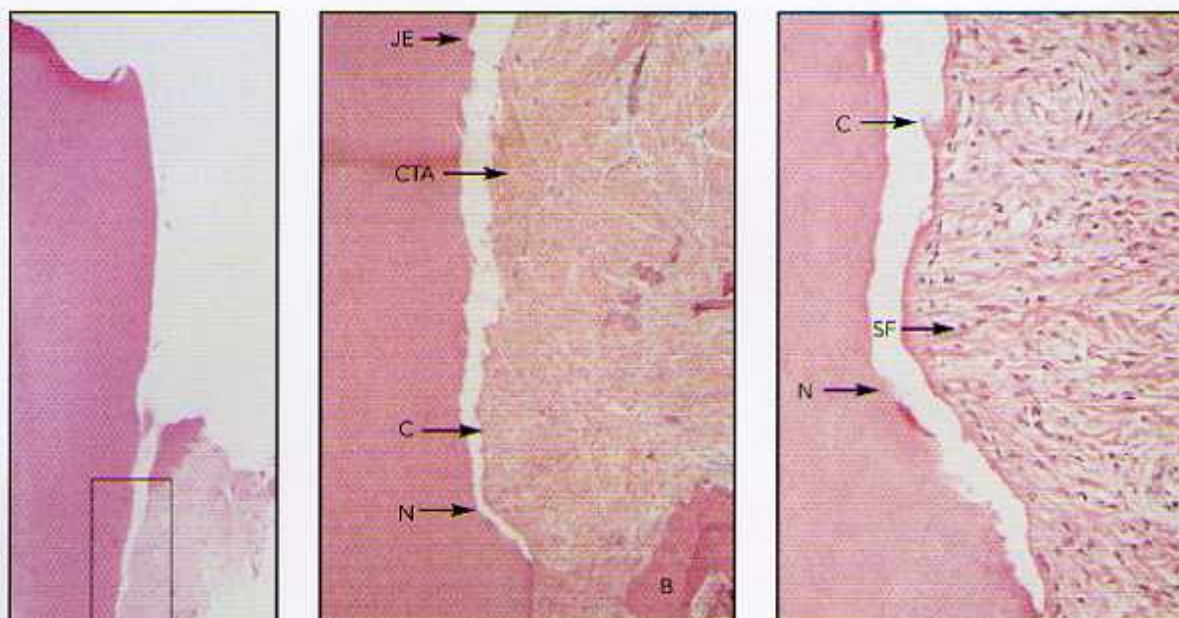
**Fig 2** Histologic views of LANAP-treated maxillary right canine from Fig 1a (hematoxylin & eosin). (left) Low-power view ( $\times 1$ ) with box around area of interest. (center) Medium-power view ( $\times 16$ ) showing calculus notch (N) with new cementum (C) in and coronal to the notch and old cementum (OC) apical to the notch, apical extent of junctional epithelium (JE), and new bone (B) adjacent to the notch. (right) High-power view ( $\times 40$ ) of notch area demonstrating new cementum (C) filling the notch (N) and extending coronally, old cementum apical to the notch (OC) covered by new cementum, new alveolar bone (B), and new periodontal ligament and gingival fibers attached to the tooth.

ontrol teeth. Clinical results are presented in Table 1. mGI, PI, and BOP were improved on all test and control teeth. Total energy applied to the test pock-

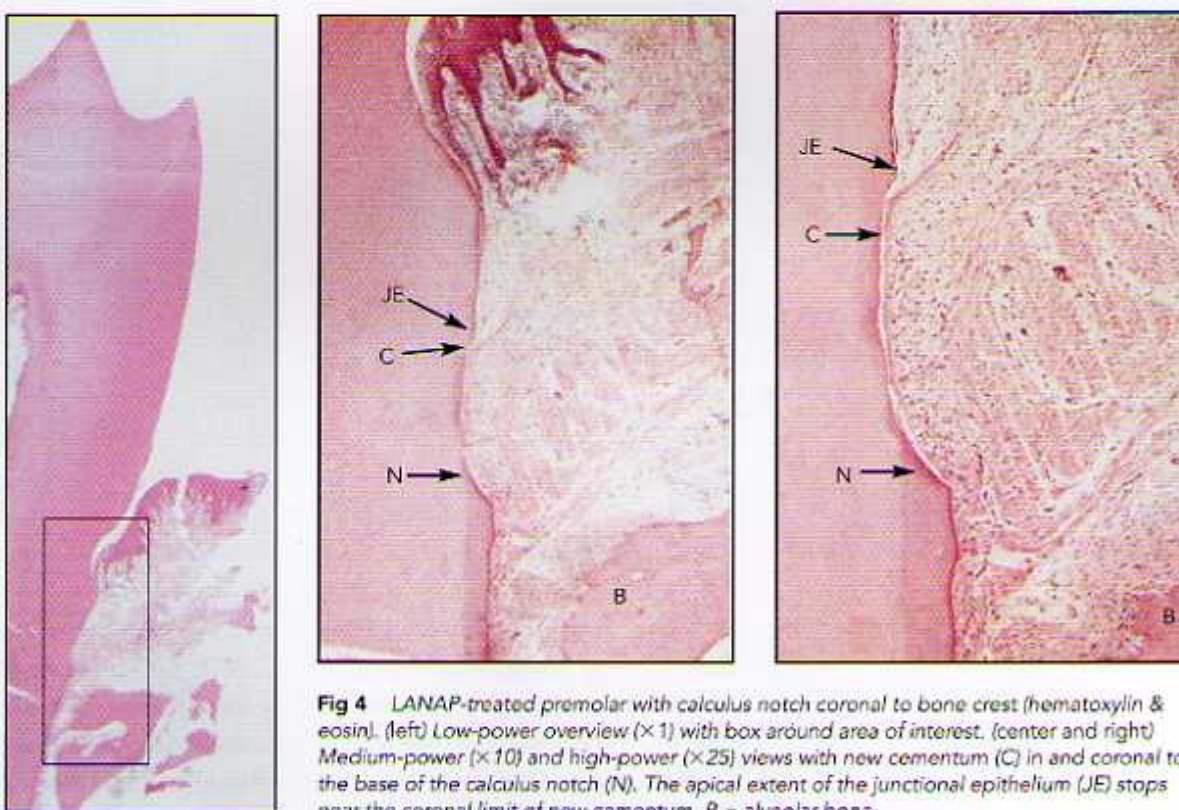
ets ranged from 14 to 25 J/mm of probing depth (mean 19 J/mm).

All six LANAP-treated specimens showed new CEM and new CTA in and

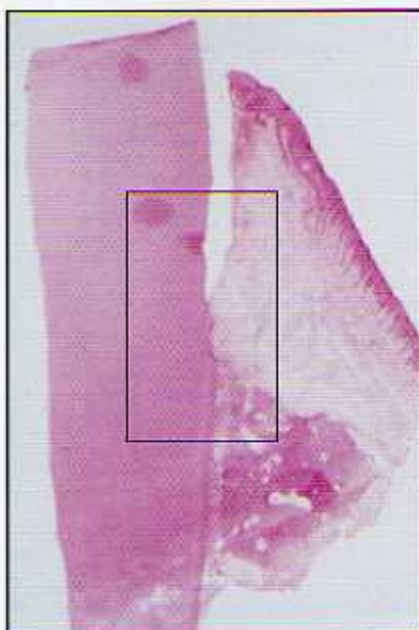
occasionally coronal to the notch (Figs 2 to 5). In two specimens, the notch was within the infrabony pocket (sub-crestal) and the new CEM and new



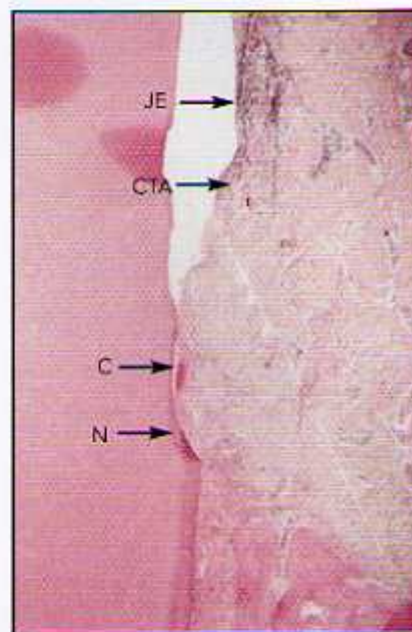
**Fig 3** LANAP-treated mandibular left second premolar of a 48-year-old man with an infrabony defect (hematoxylin & eosin). (left) Low-power view ( $\times 1$ ) outlining the area of interest. (center and right) Medium-power ( $\times 16$ ) and high-power ( $\times 63$ ) views showing the calculus notch (N), thin layer of new cementum (C) in and coronal to the base of the notch, junctional epithelium (JE) at the coronal level, new CTA with Sharpey fibers (SF), and new bone (B) adjacent to the notch. (Cementum is artificially separated from tooth.)



**Fig 4** LANAP-treated premolar with calculus notch coronal to bone crest (hematoxylin & eosin). (left) Low-power overview ( $\times 1$ ) with box around area of interest. (center and right) Medium-power ( $\times 10$ ) and high-power ( $\times 25$ ) views with new cementum (C) in and coronal to the base of the calculus notch (N). The apical extent of the junctional epithelium (JE) stops near the coronal limit of new cementum. B = alveolar bone.



**Fig 5** Canine tooth with calculus notch coronal to bone crest treated with LANAP (hematoxylin & eosin). (left) Low-power ( $\times 1$ ) view with box around area of interest. (right) Medium-power ( $\times 10$ ) view showing new cementum (C) in and coronal to the calculus notch (N). New CTA is evident between new cementum and the apical extent of the junctional epithelium (JE).



**Table 2** Frequency of histologic findings following the use of LANAP or scaling and root planing alone (3-month results, n = 6 teeth for each treatment)

Measurement/Treatment	Frequency
New cementum	
LANAP	6/6 (1.2*)
SCL/RP	1/6 (0.1*)
New bone	
LANAP	4/6
SCL/RP	2/6
New CTA	
LANAP	6/6
SCL/RP	1/6

\*Mean amount, in millimeters, from micrometer readings.

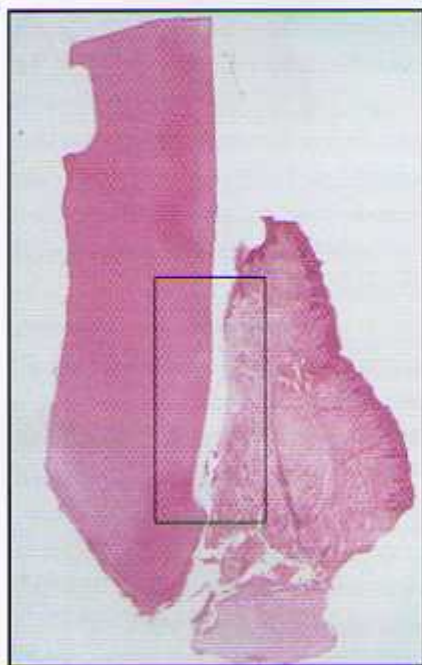
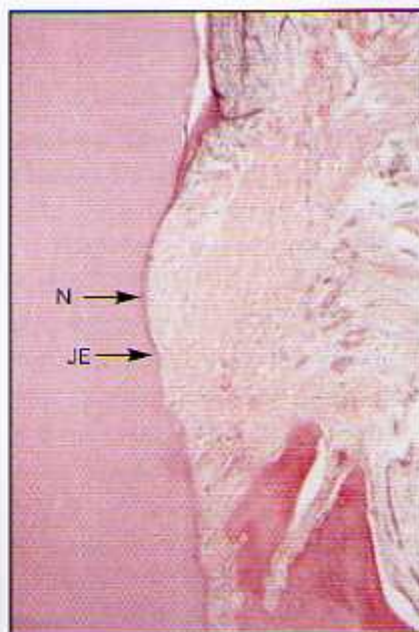
LANAP = laser-assisted new attachment procedure; SCL/RP = scaling and root planing; CTA = connective tissue attachment.

CTA were adjacent to new AB, technically showing periodontal regeneration. Five of the six control teeth had an LJE, with no evidence of new attachment or regeneration (Figs 6 and 7). One control specimen did show a small

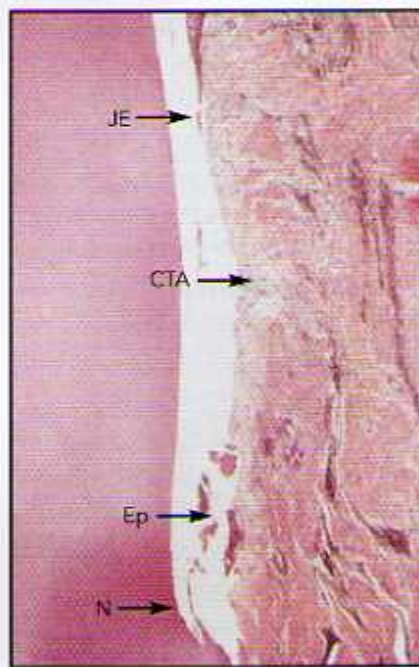
amount (0.1 mm) of new CEM and CTA. There was no evidence of any adverse histologic changes to the root surface or the pulp of any of the teeth. Histologic results are presented in Table 2.



**Fig 6** Control tooth treated with scaling and root planing without laser application (hematoxylin & eosin). (left) Low-power ( $\times 1$ ) view with box around area of interest. (right) Medium-power ( $\times 10$ ) image showing calculus notch (N) with no evidence of new CEM, new AB, or new CTA. The junctional epithelium (JE) extends to the apical extent of the notch.



**Fig 7** Mandibular premolar that received control treatment (scaling and root planing alone) (hematoxylin & eosin). (left) Low-power ( $\times 1$ ) view with box around area of interest. (right) Medium-power ( $\times 10$ ) view demonstrating lack of good tissue contact with root, even though CTA is present between the calculus notch (N) and junctional epithelium (JE). No new CEM is evident, and some epithelial islands (Ep) are present at the depth of the pocket and as islands within the connective tissue.



## Discussion

This human histologic report demonstrates favorable histologic healing with the use of the free-running pulsed Nd:YAG laser used in a specific patented technique of LANAP. Apparent periodontal regeneration (CEM, PDL, AB) on a calculus- and plaque-contaminated area of the root was seen on two of the test teeth, and CEM-mediated new attachment was evident on the other four laser-treated teeth. Similar periodontal healing in humans has been shown with other surgical techniques.<sup>2-21,58-60</sup>

The histologic assessment was based primarily on presence/absence criteria but also included linear measurements of new CEM length. As with the control teeth in this report, the literature demonstrates consistent and almost universal healing by LJE following scaling and root planing, gingival curettage, and open flap debridement procedures<sup>2,3,56,57,59-62</sup> and variable histologic results with bone replacement graft materials and miscellaneous regenerative agents on contaminated root surfaces.<sup>3-21,59,60,61,63-68</sup>

Comparison of the results of this study with those from a study that used demineralized freeze-dried bone allografts<sup>2</sup> suggests essentially equivalent histologic results using the LANAP procedure. In this study new CEM was seen in 100% of the cases versus 77% of the cases in the Bowers et al<sup>2</sup> study; new CEM length was the same (1.2 mm) as in Bowers et al; and the frequency of new CTA was 100% versus 68% for Bowers et al. It should be noted that the number of specimens was larger in the study of Bowers et al.

Treatment allocation could not be concealed from the therapist, as he had to use the laser on one tooth and not the other. Treatment (laser or no laser) was allocated according to a random code after all preliminary measurements and procedures, including placement of the calculus notch, had been completed.

Accurate placement and evaluation of the calculus notch presented several challenges. Since no flaps were reflected, direct visualization of the calculus was not possible. Positioning of the notch was based on repeated measurements from the CEJ to the clinically detectable calculus and evaluation of calculus when it was evident on the radiographs. The appropriate "depth" was marked on the shank of the quarter-round bur, and the notch was placed as carefully as possible. Again, because no flap was reflected, the depth of the notch into the root was confined to the lateral part of the bur head and was necessarily limited. It is true that there is no direct way to guarantee that the notch was actually placed in calculus, but this was the most tedious and difficult part of the entire procedure because the depth of the bur placement and therefore the depth of the notch was based on clinical and/or radiographic detection of calculus. Since clinical and radiographic detection of calculus leads to many false-negative but no false-positive results, it is felt that this was as accurate as could be accomplished with the closed procedure employed. In addition, if any error was made it was to place the notch more coronally to be sure that it was in calculus and/or contaminated root surface. On the his-

tologic slides, the position of the notch was verified by using the clinical measurements related to the CEJ or biopsy-related landmarks.

It should be emphasized that the LANAP is a combined therapy using a patented protocol (US patent #5,642,997) that includes several aspects: occlusal adjustment, splinting when needed, systemic and topical antibiotics, laser use for surgical pocket epithelium removal, scaling and root debridement, and laser use for tissue stabilization (welding) against the tooth surface with a fibrin clot. Use of the laser without attention to these other aspects may not yield the results reported here. It should be recognized that LANAP is a single-treatment surgical procedure. Since tissue is surgically removed from the lining of the pocket with the laser (rather than with a scalpel) and occlusal adjustment is an integral part of the protocol, it would appear that only qualified clinicians can legally perform the treatments in most locales.

In conclusion, this study demonstrated consistently positive histologic responses in periodontal pockets in humans treated with the LANAP CEM-mediated new attachment and occasionally apparent periodontal regeneration following a specific protocol with a free-running pulsed Nd:YAG laser were demonstrated.



## Acknowledgments

This study was supported by Millennium Dental Technologies, which provided the laser, training, and funding. The evaluations and conclusions made are solely those of the authors. The authors wish to acknowledge the histologic processing provided by Joanne Canale; the clinical assistance of Elizabeth Mayer, RDH, Stephanie Weil, CDA, RDH, and Susan Billiot, RDH; and the efforts of Julie Behan, RHIA, and Aubrey Quinn in preparing this manuscript.

## References

- Garrett S. Periodontal regeneration around natural teeth. *Ann Periodontol* 1996;1:621-666, 668.
- Bowers GM, Chadroff B, Carnevale R, et al. Histologic evaluation of a new attachment apparatus formation in humans. Part III. *J Periodontol* 1989;60:683-693.
- Bowers G, Chadroff B, Carnevale R, et al. Histologic evaluation of new human attachment apparatus in humans. Part I. *J Periodontol* 1989;60:664-674.
- Yukna RA, Mellonig JT. Histologic evaluation of periodontal healing in humans following regenerative therapy with enamel matrix derivative. A 10-case series. *J Periodontol* 2000;71:752-759.
- Mellonig JT. Enamel matrix derivative for periodontal reconstructive surgery: Technique and clinical and histologic case report. *Int J Periodontics Restorative Dent* 1999;19:9-19.
- Mellonig JT. Human histologic evaluation of a bovine-derived bone xenograft in the treatment of periodontal osseous defects. *Int J Periodontics Restorative Dent* 2000;20:19-29.
- Camelo M, Nevins ML, Schenk RK, Simion M, Rasperini G, Lynch SE, Nevins M. Clinical, radiographic, and histologic evaluation of human periodontal defects treated with Bio-Oss and Bio-Gide. *Int J Periodontics Restorative Dent* 1998;18:321-331.
- Sculean A, Chiantella GC, Windisch P, Donod N. Clinical and histologic evaluation of human intrabony defects treated with an enamel matrix protein derivative (Emdogain). *Int J Periodontics Restorative Dent* 2000;20:375-381.
- Yukna RA, Salinas TJ, Carr RF. Periodontal regeneration following use of ABM/P-15. A case report. *Int J Periodontics Restorative Dent* 2002;22:146-155.
- Camelo M, Nevins M, Lynch S, Schenk R, Simion M, Nevins M. Periodontal regeneration with an autogenous bone-Bio-Oss composite graft and a Bio-Gide membrane. *Int J Periodontics Restorative Dent* 2001;21:109-119.
- Windisch P, Sculean A, Klein F, et al. Comparison of clinical, radiographic, histometric measurements following treatment with guided tissue regeneration or enamel matrix proteins in human periodontal defects. *J Periodontol* 2002;73:409-417.
- Nevins ML, Camelo M, Lynch SE, Schenk RK, Nevins M. Evaluation of periodontal regeneration following grafting intra-bony defects with Bio-Oss Collagen: A human histologic report. *Int J Periodontics Restorative Dent* 2003;23:9-17.
- Sculean A, Windisch P, Keglevich T, Chiantella G, Gera I, Donos N. Clinical and histologic evaluation of human intrabony defects treated with an enamel matrix protein derivative combined with a bovine-derived xenograft. *Int J Periodontics Restorative Dent* 2003;23:47-55.
- Camelo M, Nevins M, Schenk R, Lynch S, Nevins M. Periodontal regeneration in human Class II furcations using purified recombinant human platelet-derived growth factor-BB (rhPDGF-BB) with bone allograft. *Int J Periodontics Restorative Dent* 2003;23:231-225.
- Cochran DL, Jones A, Mellonig JT, Schoolfield J, King G. Periodontal regeneration with a combination of enamel matrix proteins and autogenous bone grafting. *J Periodontol* 2003;74:1269-1281.
- Nevins M, Camelo M, Nevins ML, Schenk R, Lynch S. Periodontal regeneration in humans using recombinant human platelet-derived growth factor-BB (rhPDGF-BB) and allogenic bone. *J Periodontol* 2003;74:1282-1292.
- Hartman G, Arnold R, Mills M, Cochran D, Mellonig JT. Clinical and histologic evaluation of anorganic bovine bone collagen with or without a collagen barrier. *Int J Periodontics Restorative Dent* 2004;24:127-135.
- Sculean A, Windisch P, Chiantella G. Human histologic evaluation of an intra-bony defect treated with enamel matrix derivative, xenograft, and GTR. *Int J Periodontics Restorative Dent* 2004;24:326-333.
- Sculean A, Windisch P, Keglevich T, Gera I. Clinical and histologic evaluation of human intrabony defects treated with an enamel matrix protein derivative combined with a bioactive glass for the treatment of intrabony periodontal defects in humans. *Int J Periodontics Restorative Dent* 2005;25:139-147.
- Majzoub Z, Bobbo M, Atiyeh F, Cordoli G. Two patterns of histologic healing in intra-bony defect following treatment with enamel matrix derivative: A human case report. *Int J Periodontics Restorative Dent* 2005;25:283-294.
- Mellonig JT. Histologic and clinical evaluation of an allogenic bone matrix for the treatment of periodontal osseous defects. *Int J Periodontics Restorative Dent* 2006;26:561-569.
- Yukna RA, Bowers GM, Lawrence JJ, Fec PF Jr. A clinical study of healing in human following the excisional new attachment procedure. *J Periodontol* 1976;47:696-700.
- Yukna RA. A clinical and histologic study of healing following the excisional new attachment procedure in the monkey. *J Periodontol* 1976;47:701-709.
- Echeverria JJ, Caffesse RG. Effects of gingival curettage when performed 1 month after root instrumentation. A biometric evaluation. *J Clin Periodontol* 1983;10:277-286.
- Ramfjord SP, Caffesse RG, Morrison EC, et al. Four modalities of periodontal treatment compared over 5 years. *J Clin Periodontol* 1987;14:445-452.
- Litch JM, O'Leary TJ, Kafrawy AH. Pocket epithelium removal via crestal and sub-crestal scalloped internal bevel incisions. *Periodontol* 1984;55:142-148.

27. Cobb CM. Non-surgical pocket therapy: Mechanical. *Ann Periodontol* 1996;1: 443-490.
28. Myers TD. Lasers in dentistry: Their application in clinical practice. *J Am Dent Assoc* 1991;122:46-50.
29. Gold SI, Vilardi MA. Pulsed laser beam effects on gingiva. *J Clin Periodontol* 1994;21:391-396.
30. Ben Hatit Y, Blum R, Severin C, Maquin M, Jabro MH. The effects of a pulsed Nd:YAG laser on subgingival bacterial flora and on cementum: An in vivo study. *J Clin Laser Med Surg* 1996;14:137-143.
31. Gregg RH, McCarthy DK. Laser ENAP for periodontal bone regeneration. *Dent Today* 1998;17:88-91.
32. Gregg RH, McCarthy DK. Laser ENAP for periodontal ligament (PDL) regeneration. *Dent Today* 1998;17:86-89.
33. Neill NM, Mellonig JT. Clinical efficacy of the Nd:YAG laser for combination periodontitis therapy. *Pract Periodontics Aesthet Dent* 1997;9(suppl):1-5.
34. Mortiz A, Schoop U, Goharkhay K, et al. Treatment of periodontal pockets with diode laser. *Lasers Surg Med* 1998;22: 302-311.
35. Radvar M, MacFarlane TW, MacKenzie D, Whitters CJ, Payne AP, Kinane DF. An evaluation of the Nd:YAG laser in periodontal pocket therapy. *Br Dent J* 1996;180:57-62.
36. Liu CM, Hou LT, Wong MY, Lan WH. Comparison of Nd:YAG laser versus scaling and root planing in periodontal therapy. *J Periodontol* 1990;70:1276-1282.
37. White JM, Goodis HE, Rose CL. Use of the pulsed Nd:YAG laser for intraoral soft tissue surgery. *Lasers Surg Med* 1991;11: 455-461.
38. Cobb CM, McCawley TK, Killoy WJ. A preliminary study on the effects of the Nd:YAG laser on root surfaces and subgingival microflora in vivo. *J Periodontol* 1992;63:701-707.
39. Tewfik HM, Garnick JJ, Schuster GS, Sharawy MM. Structural and functional changes of cementum surface following exposure to a modified Nd:YAG laser. *J Periodontol* 1994;65:297-300.
40. Morlock BJ, Pippin DJ, Cobb CM, Killoy WJ, Rapley JW. The effect of Nd:YAG laser exposure on root surfaces when used as an adjunct to root planing. *J Periodontol* 1992;63:637-641.
41. Spencer P, Trylovich DJ, Cobb CM. Photoacoustic FTIR spectroscopy of lased cementum surfaces. *J Periodontol* 1992; 63:633-636.
42. Spencer P, Cobb CM, McCollum NM, Wieliczka DM. The effects of CO<sub>2</sub> laser and Nd:YAG with and without water/air surface cooling on tooth root structure: Correlation between FTIR spectroscopy and histology. *J Periodontol Res* 1996;31: 453-462.
43. Thomas D, Rapley JW, Cobb CM, Spencer P, Killoy WJ. Effects of the Nd:YAG laser and combined treatments on in vitro fibroblast attachment to root surfaces. *J Clin Periodontol* 1994;21:38-44.
44. Gopin BW, Cobb CM, Rapley JW, Killoy WJ. Histologic evaluation of soft tissue attachment to CO<sub>2</sub> laser treated root surfaces: An in vivo study. *Int J Periodontics Restorative Dent* 1997;17:317-325.
45. Trylovich DJ, Cobb CM, Pippin DJ, Spencer P, Killoy WJ. The effects of the Nd:YAG laser on in vitro fibroblast attachment to endotoxin-treated root surfaces. *J Periodontol* 1992;63:626-632.
46. Radvar M, Creanor SL, Gilmour VTH, et al. An evaluation of the effects of an Nd:YAG laser on subgingival calculus, dentine and cementum. An in vitro study. *J Clin Periodontol* 1995;22:71-77.
47. Maillet WA, Torneck CD, Friedman S. Connective tissue response to root surfaces resected with Nd:YAG laser or burs. *Oral Surg Oral Med Oral Pathol* 1996;82: 681-690.
48. Tokita Y, Sunakawa M, Suda H. Pulsed Nd:YAG laser irradiation of the tooth pulp in the cat: I. Effect of spot lasing. *Lasers Surg Med* 2000;26:398-404.
49. Sunakawa M, Tokita Y, Suda H. Pulsed Nd:YAG laser irradiation of the tooth pulp in the cat: II. Effect of scanning lasing. *Lasers Surg Med* 2000;26:477-484.

50. Krause LS, Cobb CM, Rapley JW, Killoy WJ, Spencer P. Laser irradiation of bone. I. An in vitro study concerning the effects of the CO<sub>2</sub> laser on oral mucosa and subjacent bone. *J Periodontol* 1997;68:872-880.
51. Friesen LR, Cobb CM, Rapley JW, Forgas-Brockman L, Spencer P. Laser irradiation of bone. II. Healing response following treatment by CO<sub>2</sub> and Nd:YAG lasers. *J Periodontol* 1999;70:75-83.
52. Lobene RR, Weatherford T, Ross NM, Lamm RA, Menaker L. A modified gingival index for use in clinical trials. *Clin Prev Dent* 1986;8:3-6.
53. Turesky S, Gilmore ND, Glickman I. Reduced plaque formation by the chloromethyl analogue of Vitamin C. *J Periodontol* 1970;41:41-43.
54. Miller SC. *Textbook of Periodontia*, ed 3. Philadelphia: Blakiston, 1950:125.
55. Sanz M, Newman MG, Anderson L, Matoska W, Otomo-Corgel J, Saltini C. Clinical enhancement of post-periodontal surgical therapy by a 0.12% chlorhexidine gluconate mouthrinse. *J Periodontol* 1989;60:570-576.
56. Dragoo MR. *Regeneration of the Periodontal Attachment in Humans*. Philadelphia: Lea & Febiger, 1981.
57. Bowers G, Granet M, Stevens M, et al. Histologic evaluation of new attachment in humans. A preliminary report. *J Periodontol* 1985;56:381-396.
58. Cole RT, Crigger M, Bolger G, Egelberg J, Selvig KA. Connective tissue regeneration to periodontally diseased teeth. A histological study. *J Periodontol Res* 1980;15:1-9.
59. Bowers G, Chadroff B, Carnevale R, et al. Histologic evaluation of new human attachment apparatus in humans. Part II. *J Periodontol* 1989;60:675-682.
60. Stahl SS, Froum S. Human intrabony lesion responses to debridement, porous hydroxyapatite implants and Teflon barrier membranes. 7 histologic case reports. *J Clin Periodontol* 1991;18:605-610.
61. Stahl SS, Froum SJ. Healing of human suprabony lesions treated with guided tissue regeneration and coronally anchored flaps. Case reports. *J Clin Periodontol* 1991;18:69-74.
62. Steiner SS, Crigger M, Egelberg J. Connective tissue regeneration to periodontally diseased teeth II. Histologic observations of cases following replaced flap surgery. *J Periodontol Res* 1981;16:109-116.
63. Stahl SS, Froum S. Histologic healing responses in human vertical lesions following the use of osseous allografts and barrier membranes. *J Clin Periodontol* 1991;18:149-152.
64. Shepard WK, Bahat O, Joseph CE, LoPiccolo P, Bernick S. Human clinical and histological responses to a Calcitite implant in intraosseous lesions. *Int J Periodontics Restorative Dent* 1986;6:46-63.
65. Stahl SS, Froum SJ. Histologic and clinical responses to porous hydroxyapatite implants in human periodontal defects: Three to twelve months postimplantation. *J Periodontol* 1987;58:689-695.
66. Nevins ML, Camelo M, Nevins M, et al. Human histologic evaluation of bioactive ceramic in the treatment of periodontal osseous defects. *Int J Periodontics Restorative Dent* 2000;20:459-467.
67. Harris RJ. Human histologic evaluation of a bone graft combined with GTR in the treatment of osseous dehiscence defects: A case report. *Int J Periodontics Restorative Dent* 2000;20:511-519.
68. Parodi R, Liuzzo G, Patrucco P, et al. Use of Emdogain in the treatment of deep intrabony defects: 12-month clinical results. Histologic and radiographic evaluation. *Int J Periodontics Restorative Dent* 2000;20:585-595.